# TROPANE ALKALOIDS FROM SCHIZANTHUS GRAHAMII

AURELIO SAN-MARTÍN, CECILIA LABBÉ, ORLANDO MURÓZ, MARIANO CASTILLO, MATÍAS REINA,\* GABRIEL DE LA FUENTE\* and ANTONIO GONZÁLEZ\*

Departamento de Quimica, Facultad de Ciencias, Universidad de Chile, Casilla 653, Santiago, Chile; \*Instituto de Productos Naturales Orgánicos, CSIC, Carretera La Esperanza 2, La Laguna, Tenerife, Spain

(Received 16 June 1986)

Key Word Index --Schizanthus grahamii; Solanaceae; tropane alkaloids; mesaconic, itaconic diesters; 3α-senecioyloxytropane.

Abstract—Three new tropanol dimers of mesaconic and itaconic acids and 3\(\alpha\)-senecioyloxytropane, together with other known alkaloids, were isolated from Schizanthus grahamii. The structures of the new compounds were established by spectroscopic and chemical transformations including 2-D long-range \(^{13}C^{-1}H\) correlations.

### INTRODUCTION

Previous chemical work on Schizanthus (Salpiglossideae) has shown that this genus accumulates a number of tropane-derived alkaloids and may thus be considered a typical member of the Solanaceae [1-3]. Additionally, it has been found that the peculiar botanical characteristics of Schizanthus are also paralleled by unique structural features of its alkaloids, namely, the utilization of either angelic, tiglic, senecioic or mesaconic acids as well as the synthesis of elaborated hygrine derivatives [C. Labbé and M. Castillo, unpublished results]. In this communication we wish to report the isolation and characterization of new tropane alkaloids from S. grahamii, including the dimeric tropanol diesters of mesaconic acid, 1 and 2 and of itaconic acid 3 for which we propose the names schizanthine C, D and E, respectively. The new alkaloid 3α-

senecioyloxytropane, together with tropine, (-)hygroline and (+)-pseudohygroline, already reported from S. hookeri [1], were also isolated from this species.

#### **RESULTS AND DISCUSSION**

The <sup>1</sup>H NMR data of compounds 1-4 are presented in Table 1. The signals of diagnostic value are (a) the chemical shift and multiplicity of the olefinic proton of the corresponding acid or diacids and (b) the position and multiplicities of the skeletal protons at the point of attachment of the ester moiety to the tropane nucleus. Thus, the signals at  $\delta 6.74$  and 6.72 in 1 and 2, respectively, indicated the presence of mesaconic acid, whereas the multiplet centred at  $\delta 6.04$  showed the presence of one angelic acid residue in both compounds. On the other hand, the triplet at  $\delta$ 5.09, integrating for two protons, showed that both tropane units present in 1 (C26H38N2O6) were linked to an ester group through C-3, whereas the attachment of the third ester residue at C-6 was evident by its characteristic multiplet (dd) at  $\delta$ 5.48. The mass spectrum of 1 showed the well established fragmentation pattern of 3,6-diacyloxytropane derivatives and the relative intensities of the ions at m/z 238, 222. 138 and 122 (Scheme 1) showed that the angeloyl residue was at C-6 [4, 5].

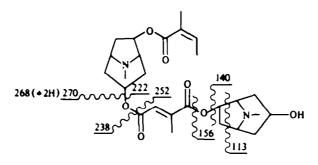
Schizanthine D (2) differed from schizanthine C (1) by an extra oxygen atom, as shown by EI mass spectrometry  $(C_{26}H_{38}N_2O_7)$ , which also showed that the angelic acid residue was at C-6 of one of the tropane units. A triplet at  $\delta 4.06$  in the <sup>1</sup>H NMR spectrum of 2 indicated a free hydroxyl group at C-3' (also corroborated by the intense ion at m/z 113, not present in 1), which together with the absorptions and multiplicities shown at  $\delta 5.69$  (H-6') 5.46 (H-6) and 5.08 (H-3) defined the substitution pattern exhibited by this alkaloid. The remaining signals in the <sup>1</sup>H NMR specta of these compounds were in agreement with the proposed structures (N-Mes, Me-mesaconic, Mes angelic, H-1 and H-5, etc.).

Schizanthine E (3), was isomeric with 2, the main differences in its  $^{1}$ H NMR spectrum being a new set of singlets at  $\delta$ 6.30, 5.72 (1H each) and 3.31 (2H) which indicated the presence of an itaconic acid residue instead

Table 1.	1H NMR data of	compounds 1-4 (20	00 MHz, CDCI	δvalues from TMS*
----------	----------------	-------------------	--------------	-------------------

Proton	1	2	3	4
3β	5.09 t	5.08 t (br)	5.01 t (br)	4.96 t (br)
	$(W_{1.2} 14)$	(4.1)	(4.5)	(5.3)
1	3.18 s (br)	$3.18 \ s \ (br)$	$3.18 \ s \ (br)$	3.09 s (br)
	$(W_{1,2},7)$	$(W_{1,2}, 9)$	$(W_{1,2}, 11)$	$(W_{1,2},7)$
5	3.32 m	3.30 m	3.28 m	
	$(W_{1/2}, 14)$	$(W_{1.2} 15)$		
6α	5.48 dd	5.46 dd	5.42 dd	
	(7.5, 3.2)	(7.5, 3.0)	(7.5, 3.0)	
7 <b>x</b>	2.54 d†	2.54 d		
	(7.5)	(7.7)		
9	6.74 q	6.72 q	3.31 s	5.60 m
	(1.5)	(1.5)		$\{W_{1,2}, 3.5\}$
11				1.96 d
				(2.0)
12	2.30 d	2.27 d	6.30 s	1.87 d
	(1.5)	(1.5)	5.72 s	(1.2)
3' B	5.09 t	4.06 t (br)	4.04 t	
	$(W_{1,2}, 14)$	(4.0)	$(W_{1\cdot 2}, 11)$	
1'		3.18 s (br)	3.18 s (br)	
		$(W_{1,2} 9)$	$(W_{1\cdot 2} 11)$	
5'	3.10 s (br)	3.30 m	3.28 m	
	$(W_{1\cdot 2} \ 10)$	$(W_{1,2}, 15)$		
6.		5.69 dd	5.68 d‡	
		(7.4, 3.0)	(7.5, 3.0)	
7' <b>β</b>		2.70 dd	2.66 dd	
		(13.6, 7.4)	(13.7, 7.5)	
10"	6.04 qq	6.04 q	6.04 q	
	(7.1, 1.4)	(7.1)	(7.0)	
11'	1.96 dq	1.97 d	1.94 d	
	(7.1, 1.4)	(7.3)	(7.3)	
12'	1.84 m	1.85 m	1.82 m	
	$(W_{1,2},5)$	$(W_{1/2}, 5)$	$(W_{1,2},5)$	
N-Mc	2.48 s	2.49 s	2.43 s	2.23 s
			2.41 s	

 $<sup>^{\</sup>bullet}J$  values in parentheses.



$$m/z$$
 $238 \xrightarrow{-C_5H_8O_2} 138$ 
 $222 \xrightarrow{-C_5H_8O_3} 122$ 
 $m/z 124$ 
 $m/z 167$ 

Scheme 1. Mass spectral fragmentation of 2.

 $<sup>\</sup>dagger dd$ , the other d overlapped by N-Me signal.  $\dagger dd$ , the other d overlapped by H-12 signal.

of mesaconic acid [6]. The gross structure of 3 was then deduced on the basis of the same reasoning as above.

The <sup>13</sup>C NMR spectra of alkaloids 1-4 (Table 2) were in agreement with the proposed structures, the assignments were made taking into account the literature data for simpler tropane alkaloids [7, 8], angelic, senecioic and mesaconic acids [9, 10] and additivity rules.

On the basis of these data alone, it was not possible to assign definitively the diester linkages to the tropane residues present in compounds 1-3. The use of partial hydrolysis to solve this problem was not attempted since the resulting half-esters would also be expected to show similar uncertainties. The specification of the mesaconic ester linkages in schizantine D (2) was resolved by application of a two dimensional long-range 13C-1H chemical shift correlation experiment. This method, which has been successfully applied in the structural determination of ingol esters [11, 12], involves 2-D longrange 13C-1H correlation using polarization transfer from skeletal and acyl protons to the ester carbonyl groups. In principle, all the carbonyl groups together with their point of attachment can be assigned simultaneously. The noise-decoupled 13C NMR spectrum of schizanthine D showed three signals in the ester carbonyl region at  $\delta$ 164.9, 166.9 and  $\overline{1}$ 67.7. The angelate carbonyl at 167.7 was readily assigned from the strong correlation observed in the contour plot of the 2-D long-range <sup>13</sup>C-<sup>1</sup>H correlation spectrum (as well as in the corresponding cross-section) with its  $\alpha$ -methyl group at  $\delta$  1.85 [9]. It also showed a weak correlation with the signal at  $\delta$ 5.46 corresponding to H-6 of the tropane nucleus, thereby confirming the previous assignment made on the basis of the mass spectral data. The C-11 carbonyl group of mesaconic ester at  $\delta$  166.9 was assigned on the basis of the strong 3-bond correlations observed with the protons at

Table 2. <sup>13</sup>C NMR data of compounds 1-6° (50 MHz, CDCl<sub>3</sub>, δ values from TMS)

C	1	2	3	4	6
	60.1	59.9	60.0	60.1	59.9
2	35.0†	34.9†	34.9†	36.7	34.41
3	67.9	67.4	67.7	66.4	67.4
4	35.6†	35.6†	35.7†	36.7	35.81
5	66.4	66.0	66.1	60.1	66.4
6	79.2	78.9	79.3	25.8	79.2
7	33.5	33.4	33.4	25.8	32.8
8	165.1	164.9	169.8	166.2	176.8
9	126.9	126.6	38.6	116.7	38.1
10	144.8	144.6	134.1	156.5	41.1
11	166.4	166.9	166.1	20.3	171.2
12	14.4	14.5	128.6	27.5	11.7
8′	167.9	167.7	167.9	-	175.2
9	128.0	127.9	128.0	-	35.9
10′	137.9	137.8	137.9		26.7
11'	15.8	15.8	15.8	-	11.7
12'	20.7	20.7	20.7		16.5
1'	59.2	59.8	60.4		59.8
2	36.7	36.01	36.6‡		35.95
3'	69.2	63.8	64.3		64.3
4'	36.7	36.91	37.4	_	37.2
5'	59.21	66.1	66.5		65.9
6'	25.9	81.3	80.8	_	80.3
7	25.9	35.9	35.8		35.8
N-Me	40.5	39.9	40.4	40.4	40.9
		40.4	40.5	_	40.1

<sup>\*</sup>DEPT multiplicity.

 $\delta 2.27$  and 6.72, assigned to H-12 and H-9, respectively [10]. The remaining carbonyl group at  $\delta 164.9$  showed a weak 4-bond correlation with the H-12 protons, together with 3-bond correlations with H-9 and with the skeletal H-3 proton at  $\delta 5.08$ , thereby establishing the attachment of the diester group in schizanthine D as depicted in 2. These and other correlations are summarized in Table 3.

Similar experiments were not performed with schizanthine E; structure 3, however, can be considered the most likely on the basis of the following observations. A comparison of the <sup>1</sup>H NMR data of tropane alkaloids esterified with  $\alpha, \beta$ -unsaturated acids with that of saturated analogues, showed that in the latter examples, H-3 absorbs normally at 0.05 · 0.1 ppm higher field than in the former. The triplet at  $\delta$ 5.01 assigned to H-3 in 3 would indicate that C-3 was attached to the unconjugated carbonyl group of the itaconic ester (cf. corresponding values in 1 and 2). This assumption was reinforced by examination of 5, the tetrahydro derivative obtained by hydrogenation of 3, which showed almost the same value for H-3, whereas H-6' was now shifted 0.07 ppm upfield. as expected. Furthermore, the physical and spectral data of 5 were identical in all respects with those of 6, also obtained by hydrogenation of 2 (see Experimental).

Basic hydrolysis of 2 and 3 gave (+)-(3R,6R)- $(tropan-3\alpha,6\beta$ -diol) [13] which defined their absolute configuration as depicted in the formulae.

3α-Senecioyloxytropane (4) was also isolated and its structure readily assigned on the basis of its spectroscopic properties and by synthesis.

<sup>†,‡</sup> Interchangeable.

Table 3. <sup>13</sup>C-<sup>1</sup>H long-range correlations in schizanthine D (2)\*

С	δ	Correlations
8′	167.7	H-12', H-6
11	166.9	H-9, H-12
8	164.9	H-12, H-9, H-3
10	144.4	H-9, H-12
10"	139.0	H-12', H-11'
9	127.6	H-12, H-11'
9	126.6	H-12, H-9 (direct)
6'	81.1	H-6' (direct), H-4', H-1'
6	78.9	H-4, H-1
3	67.3	H-5, H-1, CH2 at 1.7
3	63.8	H-5', H-1', CH <sub>2</sub> at 1.6
5	65.9	N-Me, H-1, H-3
5'	66.0	N-Me, H-1', H-3'
1'	59.9	N-Me, H-5', H-3'
1	59.8	N-Me, H-5, H-3
12'	20.5	H-12' (direct)
12	14.3	H-12 (direct), H-9

\*Experimental details of 2-D long-range of C H correlations were the same as those described in ref. [11].

#### **EXPERIMENTAL**

Schizanthus grahamii Gill was collected in Rengo, VI Region, in December. Voucher specimens are kept at the Faculty of Sciences (Universidad de Chile) Herbarium.

Isolation of alkaloids. Dried ground plant material (8.1 kg) was Soxhlet extracted with petrol and then percolated with EtOH at room temp. The EtOH extract was partitioned between acid and base in the usual manner to yield 50 g of basic material. This residue was subjected to a countercurrent distribution, in 12 funnels, between CHCl<sub>3</sub> (moving phase) and McIlvaine buffer, pH 7.1. The first five funnels contained tropine (3.3 g), (-)-hygroline (1.1 g) and (+)-pseudohygroline (1.7 g) [1]. The contents of funnels 6-9 were not analysed. The weak bases (funnels 10-12, 26 g) were partitioned again at pH 6 to yield four fractions (1.3, 1.8, 4.0 and 12 g, respectively). The third fraction was chromatographed on Al<sub>2</sub>O<sub>3</sub> (Type 60/E) and eluted with mixtures of increasing polarity of petrol and EtOAc.

Schizanthine C (1). Colourless oil (70 mg).  $[\alpha]_{25}^{25} = 17.2$  (EtOH, c 0.32). IR  $\nu_{\rm max}^{\rm fim}$  cm  $^{-1}$ : 3020, 1705, 1640, 1245.  $^{1}$ H NMR (Table 1).  $^{13}$ C NMR (Table 2). EIMS (70 eV, direct inlet) m/z (rel. int.): 474.2736 ( $C_{10}H_{38}N_2O_6$ , calc. 474.2728) [M]  $^{+}$  (14), 254.1378 ( $C_{13}H_{20}NO_4$ , calc. 254.1365) (7), 238 (2.5), 222.1523 ( $C_{13}H_{20}NO_2$ , calc. 222.1554) (17), 208.1289 ( $C_{12}H_{18}NO_2$ , calc. 208.1242) (21), 138.0928 ( $C_{8}H_{12}NO$ , calc. 138.0938) (15), 124 (100), 122 (31), 96 (38), 95 (76), 94 (94), 84 (28), 83 (39).

Schizanthine D (2). Colourless oil (243 mg).  $[\alpha]_{0.5}^{25} = 9.7$  (EtOH, c 0.584). IR  $v_{\rm max}^{\rm lim}$  cm  $^{-1}$ : 3600, 3005, 1720, 1645, 1265.  $^{1}$ H NMR (Table 1).  $^{13}$ C NMR (Table 2). EIMS m/z (rel. int.): 490.2626 ( $C_{20}H_{38}N_2O_5$ , calc. 490.2576) (7)  $[M]^*$ , 270.1363 ( $C_{13}H_{20}NO_5$ , calc. 270.1387) (2.7), 268 (3), 252.1202 ( $C_{13}H_{18}NO_4$ , calc. 252.1170) (1), 238.1409 ( $C_{13}H_{20}NO_3$ , calc. 238.1376) (4), 222.1533 ( $C_{13}H_{20}NO_2$ , calc. 222.1572) (13), 167.1044 ( $C_0H_{13}NO_2$ , calc. 167.1143) (10), 156.1075 ( $C_8H_{14}NO_2$ , calc. 156.1125) (10), 138.0920 ( $C_8H_{12}NO$ , calc. 138.0940) (21), 122 (46), 113 (80), 96 (52), 95 (86), 94 (100).

Schizanthine E (3). Colourless oil (60 mg). [ $\alpha$ ] $_{0}^{25} = 2.3$  (EtOH, c 0.086). IR  $\nu_{\max}^{61m}$  cm<sup>-1</sup>: 3660, 3002, 1715, 1705, 1640, 1245. <sup>1</sup>H NMR (Table 1). <sup>13</sup>C NMR (Table 2). EIMS m/z (rel. int.): 490.2675 ( $C_{20}H_{30}N_2O_{\gamma}$ , calc. 490.2665) (3) [M] \*, 270 (4), 268 (3), 239.1481 ( $C_{13}H_{21}NO_3$ , calc. 239.1432) (3), 222.1510 ( $C_{13}H_{20}NO_2$ , calc. 222.1527) (10), 221 (5), 156.1073 ( $C_0H_{14}NO_2$ , calc. 156.1122) (8), 140 (10), 138 (16), 122 (28), 113 (81), 96 (61), 95 (53), 94 (100).

 $3\alpha$ -Senecioyloxytropane (4). Colourless oil (287 mg). IR  $v_{\rm min}^{\rm min}$  cm  $^{-1}$ : 3005, 1700, 1650, 1040.  $^{-1}$ H NMR (Table 1).  $^{-13}$ C NMR (Table 2). EIMS m/z (rel. int.): 223.1529 ( $C_{13}H_{21}NO_2$ , calc. 223.1498) (5) [M] $^+$ , 222 (11), 208 (5), 140.1059 ( $C_8H_{13}NO_2$ , calc. 140.1043) (13), 124.1117 ( $C_8H_{14}N$ , calc. 124.1110) (100), 123 (12), 122 (16), 111 (18), 110 (26), 96 (33), 94 (55). Esterification of 80 mg of tropine with 0.8 ml of senecioyl chloride (reflux, 3 hr) gave 45 mg of a product identical in all respects to compound 4.

Tetrahydroschizanthine E (5). Obtained by hydrogenation of 2 (30 mg), disolved in EtOAc (20 ml), over Pd-charcoal (10%), and work up in the usual manner. Colourless oil. IR  $v_{\text{min}}^{\text{lam}}$  cm<sup>-1</sup>: 3660, 3600, 1720, 1240. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>): 60.88 (t, J = 7.3, H-11'), 1.11 (d, J = 6.9, H-12), 1.19 (d, J = 6.9, H-12'), 2.45 (s, 6H, N-Me), 3.08 (m, 2H, H-1, H-1'), 3.27 (m, 2H, H-5, H-5'), 4.03 (t, br, H-3'), 4.99 (t, br, H-3), 5.40 (m, H-6) 5.60 (m, H-6'). EIMS m/z (rel. int.): 494 (3.2) [M] \*, 272 (9), 270 (4.4), 224 (16), 140 (20), 138 (18), 122 (30), 113 (100), 96 (27), 95 (74), 94 (75).

Tetrahydroschizanthine D (6). IR, <sup>1</sup>H NMR and EIMS as 5. <sup>13</sup>C NMR (Table 2).

(+)-(3R,6R)-Tropan-3α,6β-diol. A soln of 22.1 mg of schizanthine D (2) in 4 ml of EtOH H<sub>2</sub>O containing 80 mg of Ba(OH)<sub>2</sub> was heated under reflux during 10 hr. The reaction mixture was evaporated to dryness and the solid residue sublimed (170°, 0.1 mm) affording 6.8 mg of crystalline product.  $[\alpha]_D^{2.5} + 11.02$  (EtOH, c 0.136) (lit. + 24.0) [13].

Acknowledgements This work was carried out under the sponsorship of the Convenio Universidad de Chile and Consejo Superior de Investigaciones Cientificas (CSIC, Spain). The authors are indebted to Dr. J. D. Connolly (University of Glasgow) for performing the 2-D long-range <sup>13</sup>C-<sup>1</sup>H correlations experiments and for helpful discussions. Financial support from the Departamento de Investigación y Bibliotecas (Universidad de Chile), Organización de los Estados Americanos and Comisión Asesora de Investigación Científica y Tècnica (Spain) is gratefully acknowledged.

## REFERENCES

- San-Martin, A., Rovirosa, J., Gambaro, V. and Castillo, M. (1980) Phytochemistry 19, 1007.
- Gambaro, V., Labbé, C. and Castillo, M. (1983) Phytochemistry 22, 1838.
- 3. Ripperger, H. (1979) Phytochemistry 18, 717.
- 4. Evans, W. C. and Major, J. A. (1966) J. Chem. Soc. C 1621.
- Evans, W. C. and Wolley, V. A. (1978) Phytochemistry 17, 171 and references cited therein.
- Pouchert, Ch. J. and Campbell, J. R. (1974) The Aldrich Library of NMR Spectra 2, 162A.
- Simeral, L. and Maciel, G. E. (1974) Org. Magn. Res. 6, 226 and references cited therein.
- Stenberg, V. I., Narain, N. K. and Singh, S. P. (1977) J. Heterocyclic Chem. 14, 225.
- Joseph-Nathan, P., Wesener, J. R. and Günther, H. (1984) Org. Magn. Res. 22, 190.
- Lippmas, E., Peck, T., Andersen, K. and Rappe, C. (1970) Org. Magn. Res. 2, 109.
- Connolly, J. D., Fakunle, C. O. and Rycroft, D. S. (1984) Tetrahedron Letters, 3773.
- Connolly, J. D., Fakunle, C. O. and Rycroft, D. S. (1984) J. Chem. Res. 368.
- 13. Fodor, G. and Kovacs, O. (1973) J. Chem. Soc. 2341.